Development and Validation of High Performance Liquid Chromatography method for analysis of Gatifloxacin & its Impurity

Narendra M.Petha, J.G.Patil, J.G.Chandorkar Indofil Industries Ltd, Gujarat *jchandorkar-icc@modi.com*

ABSTRACT

Gatifloxacin is a antibacterial agent, Rapid, sensitive and selective analytical method is essential for monitoring the different reactions steps involved in process development of Gatifloxacin. A simple isocratic reverse phase High Performance Liquid Chromatographic [HPLC] method was developed for simultaneous separation of different intermediates and other impurities. The method was utilized successfully in analyzing the reaction streams, related substances in final product and for the assay in drug.

Keywords: Gatifloxacin, Reverse phase HPLC, Impurity, Methyl Piperazine.

INTRODUCTION

Impurity profiling is the common name of a group of analytical activities, the aim of which is the detection, identification/structure elucidation and quantitative determination of organic and inorganic impurities, as well as residual solvents in bulk drugs and pharmaceutical formulations. Since this is the best way to characterize the guality and stability of bulk drugs and pharmaceutical formulations, this is the core activity in modern drug analysis. Due to the rapid development of the very analytical methodologies available for this purpose and the similarly rapid increase of the demands as regards the purity of drugs it is an important task to give a summary of the problems and the various possibilities offered by modern analytical chemistry for their solution.

Gatifloxacin level in biological fluids, in different pharmaceutical formulations and as a raw material for related substances have previously been determined spectrophotometric, by gas chromatographic [HPLC] techniques. Literature surveys reveal hardly any method for the analysis of reaction mixture obtained in the preparation of Gatifloxacin. There is an increasing need for rapid and sensitive method for the determination of raw materials, intermediates and finished products in reaction stream during process development of Gatifloxacin. The HPLC system consisted of Jasco make UV/VIS detector model 1575, along with Borwin software (Integrator) were used. Analysis were performed on Stainless steel column containing C-18 packing, 5 µ ODS [25 cm x 4.6 mm].

EXPERIMENTAL

CHROMATOGRAPHIC CONDITION: -

1) COLUMN: 250 mm x 4.6 mm i.e. - Stainless steel column containing C18, 5 μ

- 2) MOBILE PHASE: BUFFER: ACETONITRILE (80: 20)
- 3) FLOW RATE: 1.0 ml / minute.
- 4) DETECTOR WAVELENGTH: 210 nm.
- 5) SAMPLE SIZE: 20 µl.

The approximate Retention Times that should be obtained using these chromatographic conditions are:

- GATIFLOXACIN approximate retention time = 1.85 Minutes

- 2-Methyl Piperazine approximate retention time = 04.46 Minutes.

PREPARATION OF MOBILE PHASE:

Buffer [0.025 gm Ortho Phosphoric acid] 80 v & 20 v of Acetonitrile. Adjust pH 3.0 by TEA.

PREPARATION OF THE TEST SOLUTION FOR ANALYSIS OF GATIFLOXACIN:

Amber glassware must be used when preparing these solutions because GATIFLOXACIN is Photosensitive.

ANALYTICAL STANDARD SOLUTION

[A] Accurately weigh 100 mg [\pm 5 mg] of Working standard of GATIFLOXACIN & transfer to a 50 ml volumetric flask. Add 10 ml of mobile phase &

sonicate until dissolved. Allow to cool to room temperature & dilute to volume with mobile phase. [B] Accurately weigh 5 mg $[\pm 1 \text{ mg}]$ of the working standard of 2-Methyl Piperazine & transfer to 50 ml volumetric flask. Add 10 ml of mobile phase & dilute to volume with mobile phase.

[C] Transfer 4.0 ml of solution [B], in solution [A] in volumetric flask. & Dilute to volume with Mobile phase & mix thoroughly.

PREPARATION OF SAMPLE SOLUTION:

Accurately weigh 100 mg of sample & transfer to 50 ml volumetric flask. Add 10 ml of Mobile phase & sonicate until dissolved. Allow to cool to room temperature & dilute to volume with mobile phase.

INJECTION PRECISION:

Make duplicate injections of the analytical standard solution. Using a computing Integrator measure the GATIFLOXACIN peak area in the injections made.

The relative Standard deviation must not be greater than + 2.0%.

CALCULATION OF RESULTS: 1. GATIFLOXACIN CONTENT:

 $\begin{array}{rl} & & A_{\text{Samp.}} & x \ W_{\text{Std.}} \\ \text{GATIFLOXACIN Content} & = & & & \\ & & A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \\ \end{array} \\ \end{array} \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Samp.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right. \\ \left. \begin{array}{c} A_{\text{Std.}} & x \ W_{\text{Std.}} \end{array} \right.$

A_{samp} = Area of GATIFLOXACIN peak in an injection of sample.

A_{std.} = Mean Area of GATIFLOXACIN Peak in injection of analytical standard solution.

W_{Samp.} = Weight of the sample taken to prepare relevant sample solution. (in mg)

 $W_{Std.}$ = Weight of GATIFLOXACIN reference standard taken to prepare analytical standard Solution. (in mg) P = Known Purity of GATIFLOXACIN Reference standard.

2. 2-Methyl Piperazine CONTENT:

2-Methyl Piperazine Content = A_{Samp.} x W_{Std.} A_{Std.} x W_{Samp.}

 $A_{\text{samp.}}$ = Area of 2-Methyl Piperazine peak in an injection of sample.

 A_{std} = Mean Area of 2-Methyl Piperazine peak in injections of analytical standard solution.

 $W_{Samp.}$ = Weight of the sample taken to prepare relevant sample solution. (in mg)

 W_{Std} = Weight of 2-Methyl Piperazine reference standard taken to prepare analytical standard solution. (in mg)

P = Known Purity of 2-Methyl Piperazine reference standard.

RESULTS AND DISCUSSIONS

1. SYSTEM SUITABILITY:

System suitability data as shown in Table No. 1 shows method is accurate.

Table No. 1: -

Compound	Standard Deviation	RSD	Theoretical Plates	Resolution Factor	Tailing Factor
2-METHYL PIPERAZIN	5.04619	0.6621	5370	1.6	1.1
GATIFLOXACIN	302.6325	1.624284	7271	-	-

2.SUPPORTING DATA FOR TEST PROCEDURE REPRODUCIBILITY IN ASSAY TEST

Data in Table No. 2 shows method is rugged & reproducible.

Table No. 2

GATIFL	OXACIN			2-Methyl Pipe	erazine	
DAY	WEIGHT	Х	DAY	WEIGHT		Х
	[mg/ml]	[%]		[mg/ml]		[%]
28/09/2004	1	99.32	28/09/2004	0.01		98.73
		98.84				99.12
		99.5				99.36

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29/09/2004	1			29/09/2004	0.01	
			99.57			98.83
			99.24			99.21
			99.14			99.08
30/09/2004	1			30/09/2004	0.01	
			99.38			99.33
			99.72			98.54
			99.71			99.34
	AVG. [%]		99.38			99.06
	S.D.	(0.2844732			0.29580399
	RSD	(0.2862479			0.29861093
	2V	(0.5724959			0.59722186

3.RECOVERY-

Data in Table No. 3 shows method has recovery more than 99% so method is rugged and accurate. **Table No. 3**

	OF GATTIFLOXACINE				SUPPORT	NG DATA	
					FOR TEST P	ROCEDURE	
					ACCURACY		
	Concentration	A (mg/ml)		B (mg/ml)		x	
	[STD]	-		0.0770		0.0770	
	1.0 [mg/ml]	1		0.9778		0.9778	
		1		0.9847		0.9847	
		1		0.9846		0.9846	
		1		1.0031		1.0031	
			Х	0.98755			
			S	0.01086			
			CV	1.09949			
100% of The	eoretical Concentration 9	95% Confidence		0.98755	0.01726426		
Limits =		1					
Limits =							
	DF 2-METHYL PIPERAZIN				SUPPORT	NG DATA	
	DF 2-METHYL PIPERAZIN						
	DF 2-METHYL PIPERAZIN				SUPPORT		
	Concentration	A (mg/ml)		B (mg/ml)	SUPPORT FOR TEST P		
	Concentration [STD]				SUPPORT FOR TEST P	ROCEDURE	
	Concentration	A (mg/ml)		B (mg/ml) 0.00993	SUPPORT FOR TEST P	ROCEDURE I X 0.9925	
	Concentration [STD]	A (mg/ml) 0.01 0.01		B (mg/ml) 0.00993 0.00995	SUPPORT FOR TEST P	ROCEDURE Image: Constraint of the second secon	
	Concentration [STD]	A (mg/ml) 0.01 0.01 0.01		B (mg/ml) 0.00993 0.00995 0.00991	SUPPORT FOR TEST P	ROCEDURE I X I 0.9925 I 0.9946 I 0.9905 I	
	Concentration [STD]	A (mg/ml) 0.01 0.01		B (mg/ml) 0.00993 0.00995	SUPPORT FOR TEST P	ROCEDURE Image: Constraint of the second secon	

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4	0

			Х	0.99594		
			S	0.00701		
			CV	0.70377		
100% of T	heoretical Concentration	n 95% Confidence		0.99594	0.01114444	
	Limits =					

A- Actual Concentration Taken.

B- Actual CONCENTRATION Recover.

C-X-Recovery.

4. SUPPORTING DATA FOR TEST PROCEDURE ACCURACY

Data in Table No 4.1 and 4.2 show the linearity curve (Slope) & regression data for the product and its impurity, which confirms method, is accurate & reproducible.

Table No. 4.1

ASSAY OF GATIFLOXACIN	SUPPORTING DATA FOR TEST
	PROCEDURE ACCURACY

CRITERIA MEASURED	DRUG SUBSTANCE ACCEPTANCE VALUE	RESULTS
Concentration Range	40 – 150%	-
Graphic Plot	24 Points	
R		0.99837
R^2		0.996743
Average Fractional Recovery [X] (Average of x X 100)		
i.e. =6	99 – 101%	100.06%
Slope (A Vs B)	1.00 ideal	1.014
Cv= (i.e. Average of all Cv)	-	0.7730688

CV = Coefficient of Variation

Table No. 4.2

CONTENT OF RELATED SUBSTANCE –	SUPPORTING DATA FOR TEST
2-Methyl Piperazine	PROCEDURE ACCURACY

CRITERIA MEASURED	DRUG SUBSTANCE ACCEPTANCE VALUE	RESULTS
Concentration Range	40 – 150%	-
Graphic Plot	24 Points	
R		0.99994
R ²		0.99988
Average Fractional Recovery [X] (Average of x X 100)		
i.e =	95 – 105%	99.20%
6		
Slope (A vs B)	1.00 ideal	0.989
Cv= (i.e. Average of all Cv)	-	0.72090

RESULTS AND DISCUSSIONS

As per USP XXVII, system suitability was carried out freshly prepared reference solution B to check various parameters such as efficiency, resolution and peak tailing which found to comply with USP requirements. (Refer Table No1)

The content of an impurity in Gatifloxacin by proposed method. The lower values of reproducibility indicate that the method is precise and accurate. The mean recoveries of Impurity were in the range of 99.3% to 100%, which shows that there is no interference from the mobile phase, which also confirm the reproducibility and reliability of the method.

CONCLUSION

i)The proposed method is simple rapid and selective.

ii) Percent Relative standard Deviation was very slow, below 2.0% which indicate that method id highly precise & reproducible.

iii)Short Analysis time (less than 10Min)coupled with simplicity and ease of operation warrants use of the method for analysis of Gatifloxacin along with its impurity stated above in Bulk and well as in Formulated dosages for Assays and for said Related Substance by HPLC.

iv)Therefore, method can be useful in routine quality Control analysis in bulk

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